

Atty Docket No. PC19461A
Appl. No. 10/521,336
Reply to Office action of 11/15/2006

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This listing of claims will replace all prior versions, and listings, of claims in the application:

Listing of Claims:

1.-23. (canceled)

24. (currently amended) Antibiotic 107891, isolated from Microbispora sp. ATCC PTA-5024 complex comprising Factor A1 and Factor A2 being a white powder having the following characteristics:

(A) Mass spectrum recorded from a 0.2 mg/ml solution in methanol:water 80/20 (v/v) with trifluoracetic acid 0.1% on a ThermoFinnigan LCQ Deca instrument fitted with an electrospray source, using ThermoFinnigan calibration mix under the following electrospray conditions: spray voltage: 4.7 kV; capillary temperature: 220.degree. C.; capillary voltage: 3V; infusion mode 10 .mu.l/min, showing two double protonated ions at m/z 1124 and m/z 1116, corresponding to the lowest isotope composition of Factor A1 and A2, respectively;

(B) Infrared spectrum recorded in KBr with a Bruker FT-IR spectrophotometer model IFS 48, exhibiting absorption maxima at (cm⁻¹) 3263; 2929; 1661; 1533; 1402; 1114; 1026;

(C) U.V. spectrum performed in methanol:H₂O 80:20 (v/v) with a Perkin-Elmer spectrophotometer Lambda 16, exhibiting two shoulders at 226 and 267 nm;

D) ¹H-NMR spectrum recorded at 600 MHz in the mixture methanol-d4:H₂O (pH 4.3 HCl) 40:10 (v/v) at 40 degrees C on a Bruker AMX 600 spectrometer applying a water suppression sequence using as internal standard the residual signal of methanol-d4 at 3.31 ppm, exhibiting the following signals [δ=ppm multiplicity; (attribution)]: 0.93 d (CH₃), 0.98 d (CH₃), 1.07 t (overlapped CH₃'s), 1.18 t (overlapped CH₃'s), 1.26 s (CH₃), 1.30 t (overlapped CH₃'s), 1.62-1.74 m (CH₂), 1.78 d (CH₃), 1.80 d (CH₃), 2.03 m (CH₂), 2.24 m (CH), 2.36 m (CH₂), 2.72-3.8 m (peptidic alpha CH's), 3.8-5.2 m (peptidic alpha CH's), 5.53-6.08 s (CH₂), 5.62 d (CH double bond), 6.42 m (CH), 6.92 d (CH double bond), 7.0-7.55 m (aromatic CH's), 7.62-10.4 d and m (aromatic and peptidic NH's);

E) ¹³C-NMR spectrum recorded in the mixture methanol-d4:H₂O (pH 4.3 HCl) 40:10 (v/v) at 40 degrees C. on a Bruker AMX 600 spectrometer, using as internal standard the residual signal of methanol-d4 at 49.15 ppm, exhibiting the following signals: [δ=ppm; (attribution)]: 13.6-23.2 (aliphatic CH₃'s), 26.16-73 (aliphatic CH₂'s and peptidic alpha CH's), 105-136 (aromatic and double bonds CH's and quartenary carbons), 164.3-176.3 (peptidic carbonyls);

F) the acid hydrolysate in 6N HCl, (105 degrees C., 24 h) showing the presence of the following amino acids, along with other unidentified peaks, after derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate: lanthionine, methyllanthionine, glycine, proline, valine, aspartic acid (hydrolysis product of asparagine), phenylalanine and leucine;

G) the acid hydrolysate in 4N methanesulfonic acid containing 0.2% (w/v) 3-(2-aminoethyl) indole as catalyst (115 degrees C., 16 h) showing the presence of 5-chlorotryptophan; and

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H) a basic ionizable function detected by acid/base titration performed with 0.01 N potassium hydroxide in 2-methoxyethanol (MCS):H₂O 12:3 (v/v) containing a molar excess of 0.01 N hydrochloric acid.

25. (currently amended) Antibiotic 107891, isolated from *Microbispora* sp. ATCC PTA-5024 Factor A1 being a white powder having the following characteristics:

A) a doubly protonated ion at m/z 1124 corresponding to the lowest isotope composition in mass spectrum recorded from a 0.1 mg/ml solution in acetonitrile:water 50:50 (v/v) with acetic acid 0.5% on a ThermoFinnigan LCQ Deca instrument fitted with an electrospray source, using ThermoFinnigan calibration mix under the following electrospray conditions: spray voltage: 4.7 kV; capillary temperature: 250.degree. C.; capillary voltage: 8V; infusion mode 10 μ l/min;

B) the exact mass of antibiotic determined by using a Bruker Daltonics APEX II, 4.7 Tesla spectrometer fitted with an electrospray source, corresponding to a molecular weight of 2246.71 \pm 0.06, calculated monoisotopic mass from [M+2H]²⁺ at m/z 1124.36124 (accuracy 30 ppm);

C) when dissolved in CD₃CN:D₂O (1:1), ¹H NMR spectrum exhibiting the following groups of signals (in ppm) at 600 MHz using CD₃CN as internal standard (1.94 ppm), [δ =ppm, multiplicity; (attribution)]: 0.84 d (CH₃), 0.89 d (CH₃), 0.94 t (overlapped CH₃'s), 1.1 d (CH₃), 1.13 d (CH₃), 1.15 t (overlapped CH₃'s), 1.49 m (CH₂), 1.69 d (CH₃), 1.75 m (CH₂), 2.11 m (CH), 2.26 m (CH), 2.5 m (CH₂), 2.68-3.8 m (peptidic CH_β's), 3.8-5.0 m (peptidic CH_α's), 5.45-6.17 s (CH₂), 5.58 d (CH double bond), 6.36 m (CH), 6.86 d (CH double bond), 7.0-7.45 m aromatic CH's);

D) when dissolved in CD₃CN:D₂O (1:1), .sup.¹³C NMR spectrum exhibiting the following signals (in ppm) at 600 MHz using CD₃CN as internal standard (1.39 ppm), [δ =ppm; (attribution)]: 13.6-23.03 (aliphatic CH₃'s), 25.69-77.9 (aliphatic CH₂'s and peptidic α 's), 105-137.3 (aromatic and double bonds CH's and quaternary carbons), 165.6-176.6 (peptidic carbonyls);

E) Infrared spectrum recorded in KBr with a Bruker FT-IR spectrophotometer model IFS 48 exhibiting absorption maxima at (cm⁻¹): 3294; 3059; 2926; 1661; 1529; 1433; 1407; 1287; 1114; 1021;

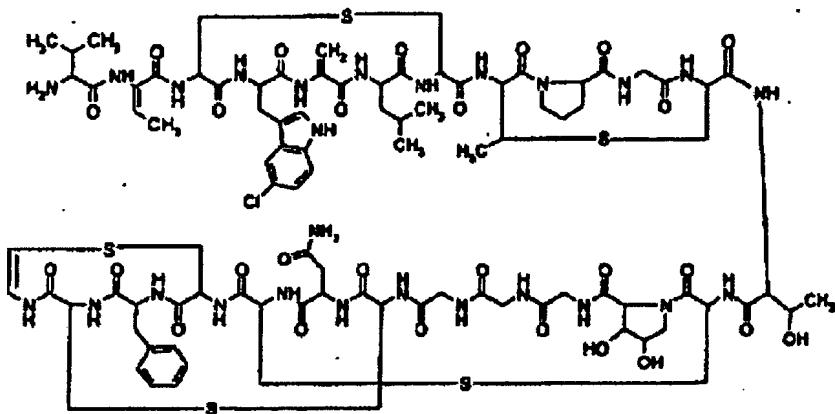
F) U.V. spectrum recorded in methanol:H₂O (in ratio 80:20) with a Perkin-Elmer spectrophotometer Lambda 16 exhibiting two shoulders at 226 and 267 nm;

G) The acid hydrolysate in 6N HCl, (105 degrees C, 24 h) showing the presence of the following amino acids, along with other unidentified peaks, after derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate: lanthionine, methyllanthionine, glycine, proline, valine, aspartic acid (hydrolysis product of asparagine), phenylalanine, and leucine; and

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H) the acid hydrolysate in 4N methanesulfonic acid containing 0.2% (w/v) 3-(2-aminoethyl)indole as catalyst (115 degrees C., 16 h) showing the presence of 5-chlorotryptophan.

26. (currently amended) The antibiotic 107891, isolated from Microbispora sp. ATCC PTA-5024 Factor A1 of claim 25, wherein the antibiotic 107891, isolated from Microbispora sp. ATCC PTA-5024 Factor A1 has the following structural formula:



27. (currently amended) Antibiotic 107891, isolated from Microbispora sp. ATCC PTA-5024 Factor A2 being a white powder having the following characteristics:

A) a doubly protonated ion at m/z 1116 corresponding to the lowest isotope composition in mass spectrum recorded from a 0.1 mg/ml solution in acetonitrile:water 50:50 (v/v) with acetic acid 0.5% on a ThermoFinnigan LCQ Deca instrument fitted with an electrospray source, using ThermoFinnigan calibration mix under the following electrospray conditions: spray voltage: 4.7 kV; capillary temperature: 250 degrees C.; capillary voltage: 8V; infusion mode 10 µl/min;

B) the exact mass determined by using a Bruker Daltonics APEX II, 4.7 Tesla spectrometer fitted with an electrospray source, corresponding to a molecular weight of 2230.71.+-.0.06, calculated monoisotopic mass from [M+2H]²⁺ at m/z 1116.36260 (accuracy 30 ppm);

(C) when dissolved in CD₃CN:D₂O (1:1), .sup.1H NMR spectrum exhibiting the following signals (in ppm) at 600 MHz using CD₃CN as internal standard (1.94 ppm), [δ =ppm, multiplicity; (attribution)]: 0.84 d (CH₃), 0.88 d (CH₃), 0.94 d (CH₃), 1.06 d (CH₃), 1.14 d (CH₃), 1.48 m (CH₂), 1.65-1.75 m (CH₂), 1.67 d (CH₃), 2.15 m (CH), 2.25 m (CH), 2.5 m (CH₂), 2.77-3.8 m (peptidic CH₂'s), 3.8-4.9 m (peptidic CH .alpha.'s), 5.45-6.14 s (CH₂), 5.59 d (CH double bond), 6.34 m (CH), 6.84 d (CH double bond), 7.0-7.42 m (aromatic CH's);

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D) when dissolved in CD₃CN:D₂O (1:1), ¹³C NMR spectrum exhibiting the following signals (in ppm) at 600 MHz using CD₃CN as internal standard (1.39 ppm), [δ=ppm; (attribution)]: 13.6-22.9 (aliphatic CH₃'s), 25.65-73 (aliphatic CH₂'s and peptidic CH_o's), 105-137.3 (aromatic and double bonds CH's and quaternary carbons), 165.7-176.1 (peptidic carbonyls);

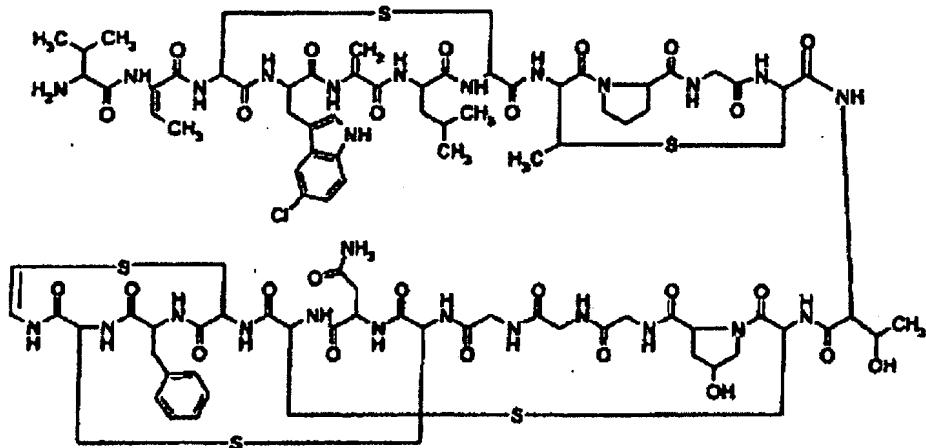
E) infrared spectrum recorded in KBr with a Bruker FT-IR spectrophotometer model IFS 48, exhibiting absorption maxima at (cm⁻¹): 3296; 3060; 2928; 1661; 1529; 1433; 1407; 1288; 1116;

F) U.V. spectrum recorded in methanol:H₂O (in ratio 80:20) with a Perkin-Elmer spectrophotometer Lambda 16 exhibiting two shoulders at 226 and 267 nm;

G) the acid hydrolysate in 6N HCl, (105° C., 24 h) showing the presence of the following amino acids, along with other unidentified peaks, after derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate: lanthionine, methyllanthionine, glycine, proline, valine, aspartic acid (hydrolysis product of asparagine), phenylalanine and leucine; and

H) the acid hydrolysate in 4N methanesulfonic acid containing 0.2% (w/v) 3-(2-aminoethyl)indole as catalyst (115°C., 16 h) showing the presence 5-chlorotryptophan.

28. (currently amended) The antibiotic 107891, isolated from *Microbispora* sp. ATCC PTA-5024 Factor A2 of claim 27, wherein the antibiotic 107891 Factor A2 has the following structural formula:



29. (currently amended) A process for producing antibiotic 107891, isolated from *Microbispora* sp. ATCC PTA-5024 and its Factors A1 and A2 and the salts thereof, comprising the steps of: cultivating *Microbispora* sp. ATCC PTA-5024 or a variant or mutant thereof maintaining the ability to produce said antibiotic, under aerobic conditions, in an aqueous nutrient medium containing an assimilable source of carbon, nitrogen and inorganic salts; isolating the resulting antibiotic from the mycelium and/or the filtered fermentation broth; and purifying the isolated antibiotic 107891.

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30. (previously presented) The process according to claim 29, wherein the strain Microbispora sp. ATCC PTA-5024 or the antibiotic 107891 producing a variant or mutant thereof are pre-cultured.

31. (previously presented) The process according to claim 29, wherein the isolation of the antibiotic 107891 is carried out by filtering the fermentation broth and the antibiotic is recovered from the filtered fermentation broth according to a technique selected from the group consisting of extraction with a water-immiscible solvent, precipitation by adding a non-solvent or by changing the pH of the solution, absorption chromatography, partition chromatography, reverse phase partition chromatography, ion exchange chromatography, molecular exclusion chromatography, and a combination of two or more of said techniques.

32. (previously presented) The process according to claim 29, wherein the isolation of the antibiotic 107891 is carried out by separating the mycelium from the supernatant of the fermentation broth and the mycelium is extracted with a water-miscible solvent whereby, after the removal of the spent mycelium, a water-miscible solution containing the crude antibiotic is obtained, which can be processed either separately or in pool with the filtered fermentation broth to recover the antibiotic 107891 by means of a technique selected from the group consisting of extraction with a solvent, precipitation by adding a non-solvent or by changing the pH of the solution, absorption chromatography, partition chromatography, reverse phase partition chromatography, ion exchange chromatography and molecular exclusion chromatography, and a combination of two or more of said techniques.

33. (previously presented) The process according to claim 32, wherein the concentration of the water-miscible solvent in the mycelium extract is reduced before it is processed to recover the antibiotic therefrom.

34. (previously presented) The process according to claim 31, wherein the filtered fermentation broth is contacted with an absorption resin, and said resin is eluted with a polar, water-miscible solvent or a mixture thereof with water, whereby a solution containing the crude antibiotic 107981 is obtained.

35. (previously presented) The process according to claim 34, wherein the absorption resin is selected from the group consisting of a polystyrene, a mixed polystyrene-divinylbenzene, and a polyamide resin.

36. (previously presented) The process according to claim 32, wherein the mycelium is extracted with a C 1-C 3 alkanol, and the mycelium extract is contacted with an absorption resin, and eluted therefrom

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with a polar water-miscible solvent or a mixture thereof with water, whereby a solution containing the crude antibiotic 107891 is obtained.

37. (previously presented) The process according to claim 36, wherein the solutions containing the crude antibiotic 107891 are pooled and processed for further purification of said antibiotic 107891.

38. (previously presented) The process according to claim 36, wherein the solution containing the crude antibiotic 107981 is concentrated and then freeze-dried to yield a crude antibiotic 107891 solid product.

39. (previously presented) The process according to claim 34, wherein the absorption resins containing the absorbed antibiotic are pooled and their mixture is eluted with a polar, water-miscible solvent or a mixture thereof with water.

40. (previously presented) The process according to claim 29, wherein the antibiotic 107981 is purified by means of a chromatographic procedure.

41. (previously presented) The process according to claim 40, wherein the chromatographic procedure is selected from the group consisting of preparative HPLC and medium pressure chromatography.

42. (previously presented) The process according to claim 29, wherein Factor A1 and Factor A2 are separated by preparative HPLC from the purified antibiotic 107891.

43. (previously presented) A pharmaceutical composition comprising an antibiotic selected from antibiotic 107891, antibiotic 107891 Factor A1, antibiotic 107891 Factor A2, and a mixture of said Factors in any proportion or a pharmaceutically acceptable salt thereof with an acid.

44. (previously presented) The pharmaceutical composition according to claim 43, further comprising a pharmaceutically acceptable carrier.